







T6PP099:

Molecular diversity in A. glabrata and prebreeding for introgressing novel resistance for stem rot and late leaf spot in cultivated groundnut

<u>Veerendra Kumar HV</u> ^{1,2}, Sudini HK¹, Sharma V¹, Bomireddy D^{1,2}, Senthil N ¹, Singh K¹, Yogendra K¹ and Pandey MK^{1*}

The perennial Arachis glabrata belongs to Rhizomatosea section with tetraploid (R2; 2n=40) genomes and is mostly known for forage, ornamental or soil conservation crop. The genotyping of 77 accessions of A. glabrata using Thermofisher's Agriseq assay containing 5000 genome-wide SNPs showed significant diversity. In parallel, significant phenotyping diversity was observed upon phenotyping these accessions for two most important diseases of the groundnut namely stem rot and late leaf spot. Among LLS and stem rot resistant sources, several accessions showed immune reaction but most of them possessed thick and rubber like leaf structure and thicker stem structure which might have served as physical barrier for these pathogens. Promising resistance sources were identified for LLS resistance (ICG 8161, ICG 8165, ICG 8166, ICG 8185, and ICG 8965) and stem rot resistance (ICG 8185, ICG 8167, ICG 8167, ICG 8167 and ICG 8924) for potential use in pre-breeding. Interestingly, accession ICG 8168 has demonstrated resistance for both the diseases along with leaf structure similar to cultivated groundnut, therefore, we have initiated pre-breeding research for the introgression of resistance segments into elite background of high oleic genotype ICGV181008. Further research is required to understand the genetics and molecular mechanism of these resistance sources in addition to intensifying efforts in introgressing these segments in the genetic background of elite varieties.

T6PP100:

Genetic variation in CaTIFY4b improves adaptation to terminal drought conditions in chickpea

<u>Barmukh R</u>^{1,2}, Roorkiwal M¹, Garg V^{1,3}, Khan AW¹, German L⁴, Jaganathan D¹, Chitikineni A^{1,3}, Kholova J¹, Kudapa H¹, Sivasakthi K¹, Samineni S¹, Kale SM¹, Gaur PM¹, Sagurthi SR², Benitez-Alfonso Y⁴ and Varshney RK^{1,3*}

Chickpea (Cicer arietinum) is a major cool-season legume crop typically cultivated on residual soil moisture during the post-rainy season, and it is often exposed to a late-season water deficit towards the end of the crop cycle. Identifying the genetic components underlying adaptation to terminal drought stress is important for improving chickpea productivity. Here, we present the fine mapping and functional characterization of "QTL-hotspot", a genomic region on linkage group 04 (CaLGO4) in chickpea that harbours major-effect quantitative trait loci (QTLs) for key drought-adaptive traits. We report that a non-synonymous mutation in the plant-specific transcription factor CaTIFY4b regulates seed weight and organ size, and contributes to drought adaptation in chickpea. Ectopic expression of CaTIFY4b in the model legume Medicago truncatula increased root growth and architecture under water deficit. Gene expression analysis indicated that CaTIFY4b might control organ size under drought in chickpea by regulating the expression of GRF-INTERACTING FACTOR1 (GIF1), a transcriptional co-activator of Growth-Regulating Factors. Phenotypic characterization of chickpea homozygous lines linked CaTIFY4b to the regulation of key physiological parameters, including pre-anthesis water use, transpiration efficiency, root architecture, and canopy development, enabling high yield performance under rainfed field conditions. Taken together, our study offers new insights into the role of CaTIFY4b and diverse molecular and physiological mechanisms underpinning chickpea growth and production under terminal drought scenarios.

 $^{^{\}mathrm{1}}$ International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Hyderabad, India

²Agricultural College, Acharya N.G Ranga Agricultural University (ANGRAU), Tirupati, India

^{*}Email: Manish.Pandey@icrisat.org

¹International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Hyderabad, India

²Department of Genetics, Osmania University, Hyderabad, India

³Murdoch's Centre for Crop & Food Innovation, State Agricultural Biotechnology Centre, Food Futures Institute, Murdoch University, Murdoch Australia

⁴Centre for Plant Science, School of Biology, University of Leeds, Leeds, UK

^{*}Email: rajeev.varshney@murdoch.edu.au